ubiquitination active sites, raising questions of how NEDD8 and ubiquitin are transfered, and how neddylation stimulates ubiquitination. Here we performed molecular dynamics simulations to address these questions. CRLs have cullin as scaffold holding two arms. One arm, substrate binding protein, binds to substrates; the other arm, Rbx protein, binds to E2. In our simulations, we observed big conformational changes on both arms. The flexible linker on the arm of Rbx1, serving as a hinge to rotate the RING domain, thus brings E2 toward substrate to shorten the 50-60 Å distance gap to a minimum of 13 Å, while the flexible linkers on the other arm of the substrate binding protein could also serve as hinges to rotate the substrate binding domain1,2 and bring substrate toward E2 thus further shorten the distance by 7~12 Å to bridge the ubiquitin transfer distance gap. The distance gap for neddylation could also be shortened due to the conformational change during the simulations. We therefore propose that a large ensemble of conformations could provide the possible conformation to bridge the distance gap for ubiquitin transfer and NEDD8 transfer, and that neddylation stimulates the ubiquitination by stimulating conformational change of CRLs and generating a larger conformational ensemble. This project is funded by NCI contract HHSN261200800001E.

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#### 143-Pos

Evolutionary Analysis of Conformational Changes in Allosteric Proteins Jouhyun Jeon<sup>1</sup>, Yoon Sup Choi<sup>2</sup>, Jae-Seong Yang<sup>2</sup>, Hyun-Jun Nam<sup>2</sup>, Sanguk Kim<sup>1,2</sup>.

<sup>1</sup>Division of Molecular and Life Science, POSTECH, Pohang, Republic of Korea, <sup>2</sup>School of Interdisciplinary Bioscience and Bioengineering, POSTECH, Pohang, Republic of Korea.

Allostery is fundamental to control numerous biological processes and coupled to the conformational rearrangements of protein structure. The changes of residue interaction networks upon ligand binding or protein-protein interactions impact protein dynamics and function. General principles on how conformational rearrangements of residues are encoded in protein sequences remain unknown. Here, we show that the residues that are evolutionary coupled with many partners mediate the conformational rearrangement of protein allostery. Highly evolutionarily coupled residues are involved in a dynamic network which participates in the smooth transition of two allosteric states; protein allostery is built up from the interaction rearrangements of these residues. We show that the evolutionary principles of protein conformational change provide the insight into the mechanisms controlling allosteric regulation and propose a new method to identify the key residues involved in the structural transition.

### 144-Pos

### The Adenylate Kinase Transition Requires Many Easy Motions, Not a Few Hard Ones

Michael D. Daily, Qiang Cui.

University of Wisconsin-Madison, Madison, WI, USA.

Conformational transitions are functionally important in many proteins. In adenylate kinase (AK), two small domains (LID and NMP) close over the larger CORE domain; the reverse (opening) motion limits catalytic turnover. Previous experiments and computations have also shown that local motions are important. Here, we hypothesize that the open/closed (O/C) transition rate depends on many low-barrier motions rather than a few high-barrier ones. To test this hypothesis, we simultaneously characterize the contributions of rigid-body (Cartesian), backbone torsional, and contact motions to the transition state (TS). O, C, and TS sub-ensembles are derived from a double-well Go simulation based on the native contacts of the O and C crystal structures. In Cartesian space, LID closes approximately two-thirds toward CORE in the TS, and NMP closes about halfway, substantially reducing rigid-body entropy. In backbone dihedral space, the TS dynamics of LID are more consistent with the higher-entropy O ensemble, while the TS dynamics of NMP, CORE, and most interdomain hinges are more consistent with the lower-entropy C ensemble. In contact space, contacts unique to C (C-contacts) with lower interresidue distances in the O state are more likely to form in the TS than those with higher distances; the CORE-LID/NMP interfaces remain sufficiently open to bind the ligand. Thus, the TS ensemble derives substantial enthalpy from C-contacts but at a relatively low entropic cost. Together with the observation that Cartesian, dihedral, and contact motions important to the transition are weakly correlated, these results strongly support our hypothesis that many degrees of freedom are important to the TS of AK. Finally, this work may complement structural analysis and protein dynamics experiments toward identifying structural features for allosteric design in proteins.

#### 145-Pos

# Functional Pathways in Proteins Are Uncovered By Strong Disorder Canan Atilgan<sup>1</sup>, Sema Ermez<sup>1</sup>, Ozlem Keskin<sup>2</sup>, Ali Rana Atilgan<sup>1</sup>.

<sup>1</sup>Sabanci University, Istanbul, Turkey, <sup>2</sup>Koc University, Istanbul, Turkey. We propose a simple and computationally inexpensive method to determine the few residues that control the communication between functionally linked regions. A protein structure is represented as a network of residues whereby edges are determined by intra-molecular contacts[1], weighted by amino-acid pair potentials[2]. The optimal path lengths that are operative under diverse perturbation scenarios are investigated for robust residue communication[3]. Pathways along which the maximum weights are minimized (strong paths) are found to be descriptive of communication during extreme events such as allosteric control and binding. This is a kinetic viewpoint whereby the rate of signal propagation is determined along paths with the lowest barrier to be surpassed.

Here, we examine 90 interacting proteins with structurally non-redundant interfaces. We study every strong path that connects the interacting proteins by recording residue pairs forming bridges between the protein-protein complexes. We then focus on those pairs that appear along the predominant fraction of these paths. Although nearly half the surface area of a protein is involved in protein-protein interactions, this approach delineates the few key contacts that control the communication between protein complexes. We compare the results from the current approach to those from computational hot spots[4]. We find that over 60% of the most used pairs correspond to a pair of hotspots and 92% of the mostly used pairs correspond to at least one hotspot on either partner protein of the complex. The results are further corroborated by experimental findings[5,6].

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#### 146-Pos

## Long Time Scale Dynamics of Molecules With Internal Rigid Fragments Sebnem G. Essiz<sup>1</sup>. Rob D. Coalson<sup>2</sup>.

<sup>1</sup>Univ. of California at San Francisco, San Francisco, CA, USA, <sup>2</sup>Univ. of Pittsburgh, Pittsburgh, PA, USA.

Due to the large number of degrees of freedom which are present in protein systems it is still a major challenge to monitor the dynamics of molecules out to the long time scales on which functionally important conformational changes occur. We have developed a rigid body Newtonian dynamics method, in which local high frequency motion of the molecule are naturally frozen out by decomposing the molecule into linked rigid bodies, thus decreasing the number of degrees of freedom monitored in the system. The factor of ca. 10 increase in the time step which comes from the elimination of high frequency motion in rigid body Newtonian dynamics can be further extended by elimination of explicit water molecules using a Langevin dynamics prescription in which friction and random force terms are added to the potential function to mimic the effect of the solvent. As a first step in this direction we have implemented a Langevin prescription for normal modes vibrational analysis of molecules with internal rigid fragments which is developed by Durand et al. [Biopolymers, 34, 759 (1994)]. Two simple illustrative examples representing signal propagation in the membrane-bound gramicidin-A dimer with two different initial conditions are presented as numerical applications for i) rigid body Newtonian dynamics (anharmonic PES) and ii) Langevin dynamics with internal rigid fragments in the harmonic field, respectively.

### 147-Pos

# Critical Assessment of the Statistical Significance of Simulated Motions in Myosin $\boldsymbol{V}$

Tharles C. David<sup>1</sup>, Christopher M. Yengo<sup>2</sup>, Donald J. Jacobs<sup>1</sup>.

<sup>1</sup>University of North Carolina at Charlotte, Charlotte, NC, USA, <sup>2</sup>Penn State, Hershey, PA, USA.

Elastic Network Models (ENM) and Principal Component Analysis (PCA) on Molecular Dynamics (MD) simulations are well-established computational methods that identify protein motions. Both methods have limitations in sampling conformational diversity. We employ an alternative method that more efficiently samples conformations by performing a rigidity analysis of myosin V using Floppy Inclusions and Rigid Substructure Topography (FIRST). We then generate trajectories of conformations using the Framework Rigidity Optimized Dynamics Algorithm (FRODA) that is approximately four orders of magnitude faster than MD. PCA analysis of the alpha carbon positions on